

SileksMagNA™ Buccal Swab DNA/RNA Isolation Kit

Catalog No KRSW100

100 DNA/RNA isolation reactions from buccal swab or scrape

Research use only

Shipping: ambient temperature

Storage: +4°C at least for one year

Kit Contents

Components (all are ready-to-use)	Volume
SamplePrep Buffer	30 ml
START Buffer	12 ml
Lysis & Binding Buffer	24 ml
SileksMagNA™ Magnetic Particles	1 ml
Wash 1 Buffer	30 ml
Wash 2 Buffer	30 ml
Wash 3 Buffer	30 ml
Final Wash Buffer	30 ml
Elution Buffer	10 ml



Related Equipment and Reagents by Sileks

Cat. No MPR001 SileksMagNA™ Magnetic Particles

Cat. No EQRM06 MagRack6 Magnetic Rack

Cat. No EQRM16 MagRack16 Magnetic Rack

Cat. No EQRM40 MagRack40 Magnetic Rack

Cat. No EQMM201 LabMix Mini 201 Mixer

Safety (more information in product material safety data sheet (MSDS))

- When working with chemicals, wear a lab coat, disposable gloves, and protective goggles. Some components may cause health damage if swallowed, breathed, in contact with eyes or skin. Follow MSDS instructions for safety.
- Avoid mixing the kit components with strong acids and alkali.
- In case of contact with eyes, rinse immediately with plenty amount of water. In case of contact with skin, immediately wash the skin with soap and plenty amount of water.

SileksMagNA™ Buccal Swab DNA/RNA Isolation Kit

Advantages

- High DNA/RNA yield and consistent results
- High purity (>1.8 OD₂₆₀/OD₂₈₀) DNA/RNA, excellent for molecular biology applications such as PCR, reverse transcription, sequencing, restrictase digestion, sequencing on chips, hybridization on chips etc.

Applications

- Rapid and efficient isolation of DNA/RNA from buccal swab and scrape
- DNA/RNA isolation from both freshly obtained or preserved samples
- The kit can be used for isolation of genomic DNA and DNA/RNA from various pathogens.

Principle

SileksMagNA™ Buccal Swab DNA/RNA Isolation Kit is designed for efficient and rapid (30 min) isolation of DNA/RNA from buccal swab or scrape using SileksMagNA™ silica-covered magnetic particles. All components of the kit are ready to use and there is no need to supply additional reagents for preparing wash buffers. Provided components are sufficient to isolate high yield DNA/RNA from 100 standard samples. Single isolation provides up to 7 µg of pure homogeneous DNA/RNA. The kit can be successfully applied for isolation of both host genomic DNA and bacterial/viral DNA/RNA. Due to low content of mRNA in buccal swab and scrape samples, they are generally not recommended for isolation of RNA for gene expression analysis.

Isolation procedure with SileksMagNA™ Buccal Swab DNA/RNA Isolation Kit consists of a few short and simple steps:

- Sample lysis: cells are destroyed and all cell components and nucleic acids are released into the solution
- Binding: nucleic acids bind to magnetic particles
- Washing: residual contaminants are washed away while pure nucleic acids remain bound to magnetic particles
- Drying: incubation at 60°C removes traces of washing buffers
- Elution: purified nucleic acids are eluted from magnetic particles

SileksMagNA™ Buccal Swab DNA/RNA Isolation Protocol

Important Notes

- SileksMagNA™ magnetic particles are particles for isolation of total nucleic acids (both DNA and RNA). To obtain only RNA after purification, use conventional DNase treatment to remove DNA. If you intend to use DNA after the purification, use conventional RNase treatment to remove RNA.
- Precipitate may form in some buffers; however, this has no effect on the quality. Warm up the buffer at 50°C to dissolve precipitate if formed. All buffers must be mixed well before use to produce a uniform suspension.
- "Mix thoroughly" in the protocol means that the solution must be mixed either by manual pipetting (20 times) or using LabMix mixer for 5 seconds on low/medium speed. "Mix well" means that the solution must be shaken 5-10 times.

Protocol

1. Add **300 µl** of well mixed **SamplePrep Buffer** to a 1.5 ml tube.
2. With a sterile cotton swab collect the buccal epithelium by gently rubbing the inner side of the donor's cheek for 20 seconds.
3. Insert the collected buccal swab into the tube with Sample Prep buffer, mix gently by manual swirling. Remove the cotton swab collector, carefully pressing it against the tube wall to retain the excess buffer in the tube.
4. **Centrifuge** the tube at **5'000 rpm** for **3 minutes**.
5. Carefully remove the supernatant.
6. Add **120 µl** of well mixed **START Buffer** to pelleted epithelial cells and mix thoroughly.
7. Incubate at room temperature for **5 minutes**. Incubation up to 15 minutes increases DNA/RNA yield.
8. In a separate tube mix the following components: **240 µl** of well mixed **Lysis & Binding Buffer** and **7 µl** of well mixed **SileksMagNA™ Magnetic Particles**. Mix thoroughly.
9. Add the prepared suspension of magnetic particles to the tube, containing prepared sample. Mix thoroughly. Incubate for **5 minutes** at room temperature, mix once or twice during incubation.
10. To collect magnetic particles with bound DNA/RNA, place the tube in a **magnetic rack** for **1 minute**. Discard the supernatant. Be careful not to disturb magnetic particles collected at the tube wall.
11. Place the tube in a **non-magnetic rack**. Add **300 µl** of well mixed **Wash 1 Buffer** and mix thoroughly until a homogeneous suspension is obtained.
12. Place the tube in a **magnetic rack** to collect the particles. Discard the supernatant.
13. Place the tube in a **non-magnetic rack**. Add **300 µl** of well mixed **Wash 2 Buffer** and mix thoroughly until a homogeneous suspension is obtained.
14. Place the tube in a **magnetic rack** to collect the particles. Discard the supernatant.
15. Place the tube in a **non-magnetic rack**. Add **300 µl** of **Wash 3 Buffer** and mix thoroughly until a homogeneous suspension is obtained.
16. Place the tube in a **magnetic rack** to collect the particles. Discard the supernatant.
17. Place the tube in a **non-magnetic rack**. Add **300 µl** of **Final Wash Buffer** and mix thoroughly.

You can stop the procedure at this stage for best DNA and RNA preservation. Long-term storage of DNA and RNA bound to magnetic particles in Final Wash Buffer is the safest option, excellent for DNA and RNA conservation or transportation purposes. To use isolated DNA and RNA after years of storage at -20°C the procedures listed below must be carried out to obtain pure DNA and RNA.

18. Place the tube in a **magnetic rack** to collect the particles. Discard the supernatant.
19. Incubate the tube in a **thermal block at 60°C for 5 minutes** to dry the pellet of magnetic particles.
20. Add **50 µl** of **Elution Buffer**. Thoroughly resuspend particles until a homogeneous suspension is obtained. If you wish to have higher DNA/RNA concentration, use 25 µl of Elution Buffer.
21. Incubate in a **thermal block at 60°C for 5 minutes**.
22. Place the tube in a **magnetic rack** to collect the particles. Transfer the supernatant containing DNA/RNA to a fresh tube. To get rid of all particles, repeat the same with the eluate: place in a **magnetic rack** and transfer supernatant into a new tube.
23. The isolated DNA/RNA mixture can be stored at -20°C or used directly.
24. DNA is stable and suitable for a long storage. If required, treat it with RNase to remove RNA and use it for subsequent applications.
25. However, if your intention is to work with RNA, avoid long storage, as RNA is prone to degradation. Use RNA for cDNA synthesis as soon as you can. Remove DNA by DNase treatment, if necessary.
26. Check the quality of DNA and RNA on appropriate agarose gel if needed.

Troubleshooting

Problem	Possible causes	Solution
Low DNA/RNA yield	The sample contains too little cells that contain DNA/RNA, or the sample is old.	Take 2x bigger sample volume for isolation, and double all following reagent volumes accordingly. Try to perform elution in smaller volume.
	Too much starting material	Use recommended amounts of samples and all buffers. Too much initial material reduces DNA/RNA yield due to suboptimal lysis, binding and wash.
	Incomplete drying of particles before adding Elution Buffer.	Increase the drying time in thermal block after removing Final Wash buffer. Check if all liquids are evaporated before elution.
	Incomplete lysis	Mix the sample more thoroughly after adding START Buffer.
	Too big volume of Elution Buffer used	Adjust optimal amount of Elution Buffer to provide required DNA/RNA concentration.
OD ₂₆₀ /OD ₂₈₀ ratio is too low	Protein contamination	Mix all buffers and all mixes during the procedure like recommended in each step. If too high DNA/RNA concentration is suspected, dilute it and measure purity once again.